

Development of Multi-Electrode Array (MEA) Assay for Phenotypic Drug Screening



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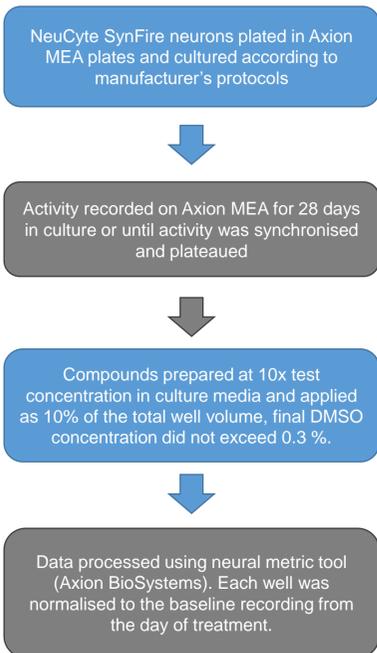
1 ABSTRACT

Multi-electrode arrays (MEA) can be utilised in drug discovery to provide a link from *in vitro* screening to *in vivo* testing, safety assays, or by modelling the functional impacts of disease mutations. Here we describe the characterisation of human iPSC derived cortical neuron co-cultures, which includes glutamatergic excitatory neurons, GABAergic inhibitory neurons, and astrocytes (NeuCyte SynFire), using the Maestro Pro (Axion BioSystems). These co-cultures were then used for compound screening and induction of seizure phenotypes.

Immunocytochemistry shows the presence of both Glutamatergic (VGLUT1) and GABAergic (GABA) neuronal markers along with astrocytic markers (GFAP). Over the course of 28 days, the neuronal co-culture shows an increase in mean firing rate as well as the development of spontaneous oscillatory activity (network bursts) with increasing synchronicity.

Diazepam (GABA_A agonist) caused a disruption of synchronous activity, decrease in mean firing rate and network burst activity, with a IC₅₀ comparable to literature data¹. Seizurogenic activity was produced in response to bicuculline (GABA_A antagonist) as shown by increase in network burst frequency and duration. These data show the suitability of human iPSC-derived neurons for compound profiling and assessment of seizurogenic liability.

2 METHODS



3 CO-CULTURE CHARACTERISATION

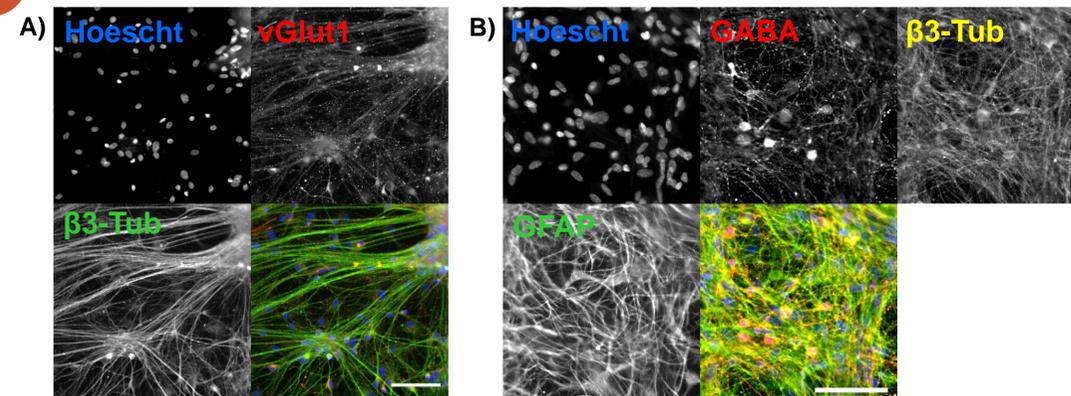


Figure 1. Human iPSC-derived neurons co-culture characterisation. The presence of individual cell types within the culture were identified by antibody staining. (A). Nuclei stain (Hoescht, blue), glutamatergic marker (vGlut1, red), and axonal marker (β3-Tub/TuJ1, green). (B). Nuclei stain (Hoescht, blue), GABAergic marker (GABA, red), and axonal marker (β3-Tub/TuJ1, yellow), and astrocyte marker (GFAP, yellow). Scale bar is 100 μm.

4 DEVELOPMENT OF NETWORK ACTIVITY

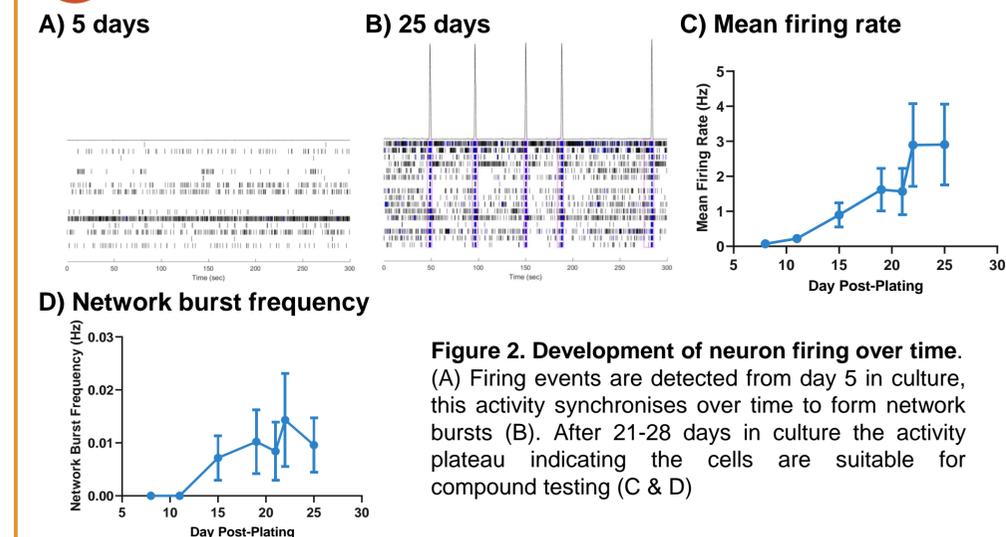


Figure 2. Development of neuron firing over time. (A) Firing events are detected from day 5 in culture, this activity synchronises over time to form network bursts (B). After 21-28 days in culture the activity plateau indicating the cells are suitable for compound testing (C & D)

5 COMPOUND PROFILING

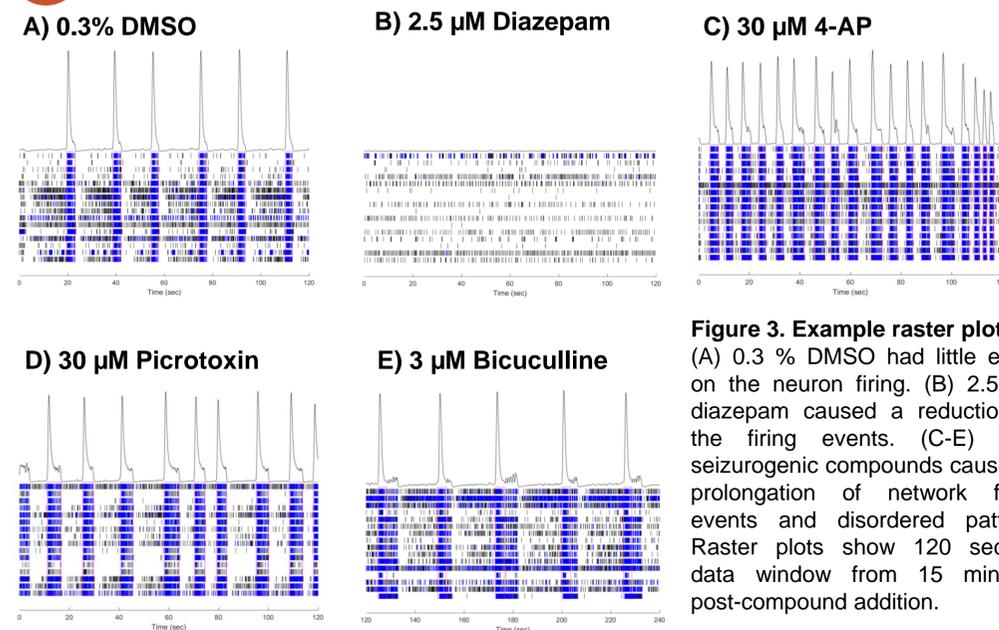


Figure 3. Example raster plots. (A) 0.3 % DMSO had little effect on the neuron firing. (B) 2.5 μM diazepam caused a reduction in the firing events. (C-E) Pro-seizurogenic compounds caused a prolongation of network firing events and disordered pattern. Raster plots show 120 second data window from 15 minutes post-compound addition.

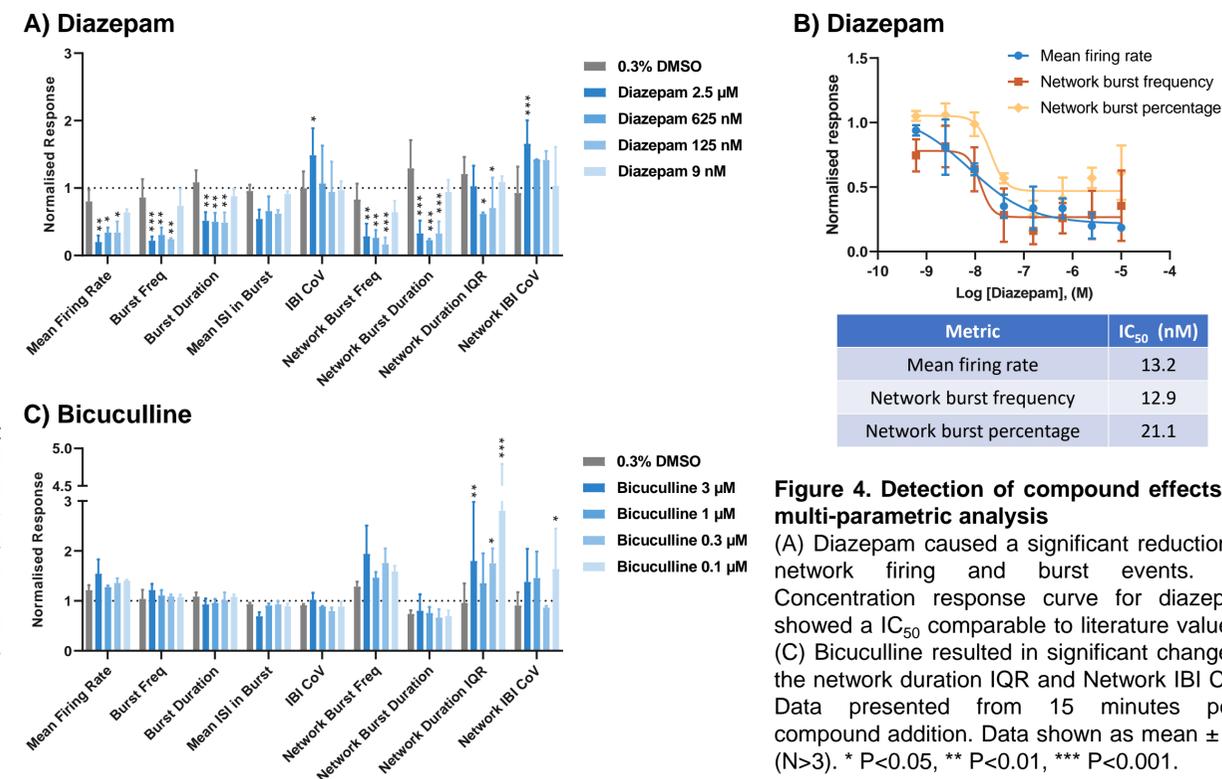


Figure 4. Detection of compound effects by multi-parametric analysis (A) Diazepam caused a significant reduction in network firing and burst events. (B) Concentration response curve for diazepam showed a IC₅₀ comparable to literature values¹. (C) Bicuculline resulted in significant change in the network duration IQR and Network IBI CoV. Data presented from 15 minutes post-compound addition. Data shown as mean ± SD (N>3). * P<0.05, ** P<0.01, *** P<0.001.

Metric	IC ₅₀ (nM)
Mean firing rate	13.2
Network burst frequency	12.9
Network burst percentage	21.1

6 CONCLUSION

- NeuCyte SynFire co-culture system including Glutamatergic, GABAergic neurons and astrocytes has been established at CRL.
- Neuronal activity monitored with Axion Maestro Pro MEA platform shows that cells display synchronised network activity from 21 *in vitro*.
- Compound profiling of known pro-seizurogenic compounds display altered network activity.
- Compound response curves can be produced. Diazepam showing similar IC₅₀ to literature data¹.

¹ Bader *et al.* PLoS One. (2017) 13;12(10):e0186147. doi: 10.1371/journal.pone.0186147.